



Ewemen Journal of Natural Product Research

Available online at http://ewemen.com/category/ejnpr/



Full Length Research

CHEMICAL CONSTITUENTS OF THE STEM OF SALACIA IMPRESSIFOLIA (MIERS) A. C. SMITH

RIPARDO FILHO H.S.¹; COSTA N.L.S.¹; PACHECO L.C.¹; ANDRADE E.S.¹; ARAÚJO R.N.M.¹; MOURÃO R.H.V.²; GUILHON G.M.S.P.¹; *SANTOS L.S.¹

¹Programa de Pós-Graduação em Química-Instituto de Ciências Exatas e Naturais, Universidade Federal do Pará-UFPA. Belém-PA, Brazil-CEP 66075-110.

²Programa de Pós-Graduação em Recursos Naturais da Amazônia and Laboratório de Bioprospecção e Biologia Experimental, Universidade Federal do Oeste do Pará, Santarém-PA, Brazil-CEP 68035-110.

ABSTRACT

Received 9 December, 2015 The plants of the genus Salacia are commonly used in the treatment of Revised on the 12 December, 2015 diabetes tipe-2, in several countries of the world. The phytochemical study Accepted 17 December, 2015 of the stem of Salacia impressifolia led to the isolation and identification of seven compounds. The compounds include the steroids sitosterol and 3-0-*Corresponding Author's Email: β -D-glucopyranosyl sitosterol, of the coumarin isoscopoletin and of the lsslouri@gmail.com triterpenes quinovic acid, 3-oxo-quinovic acid, 3-*O*-β-D-quinovopyranosyl quinovic acid, and 3-0-β-D-fucopyranosyl quinovic acid. The compounds were identified based on analysis of their NMR spectral data and comparison with literature data. This is the first report on the isolation and identification of quinovic acid and derivatives from species of the genus Salacia.

Keywords: Hippocrateaceae, *Salacia impressifolia*, quinovic acid, isoscopoletin, saponins, triterpenoids, NMR.

INTRODUCTION

Diabetes is one of the various diseases treated by the use of medicinal plants. The genus *Salacia*, with regard to treatment of diabetes type 2, has a long history in traditional medicine in Sri Lanka, India and Thailand (Tanabe *et al.*, 2009). The use of these species in the treatment of diabetes is due to the presence of potent α -glucosidase inhibitors in their constitution, for example, salacinol, kotalanol and 13 MRT (Figure 1) (Muraoka *et al.*, 2008; Oe and Ozaki, 2008). The inhibition of this enzyme promotes delayed absorption of glucose in the blood and suppresses postprandial hyperglycemia, resulting in glycemic control (Li *et al.*,

2008). Several species of this genus have been studied in search of bioactive compounds and a better understanding of their chemical composition. The genus Salacia is rich in triterpenoids belonging to several series, including ursane and oleanane obtained Salacia amplifolia et al., from (Wang 2011), bisnortriterpenes obtained from Salacia madagascariensis (Thiem al.. 2005). et quinonemethides from Salacia campestris (Carvalho et al. 2005), friedelanes and glycosides identified from Salacia chinensis (Morikawa et al., 2003; Nakamura et al., 2011), 1,3-diketofriedelane triterpene from Salacia



verrucosa (Somwong *et al.*, 2011) and lupanes obtained from *Salacia cordata* (Tinto *et al.*, 1992). This paper describes the isolation and identification of compounds **1-7** obtained from the extract of the stem bark of *Salacia impressifolia* (Miers) A. C. Smith (Hippocrateaceae).



Figure 1: Structures of salacinol, kotalanol and 13 MRT.

MATERIALS AND METHODS

Materials

NMR spectra, including ¹H-¹H COSY, DEPT and HMBC experiments, were recorded on a Varian Mercury-300 spectrometer, operating at 300 MHz at ¹H and 75 MHz at ¹³C (Varian, Palo Alto, CA, USA), CDCl₃, CD₃OD and pyridine-*d*5 were used as solvents (Sigma Aldrich, St. Louis, MO, USA). Column chromatography was performed on silica gel 60 (70–230 mesh, Macherey-Nagel, Düren, Germany).

Collection of Plant Material

The stem of *Salacia impressiflia* was collected in the Amazonas State, Brazil. A voucher specimen (No. 4321) was deposited in the Herbarium Embrapa/UFPA in Santarém, Pará State, Brazil.

Extraction and Isolation Procedure

The stem of *S. impressifolia* (2.0 kg) was air dried and extracted with MeOH at room temperature. After extraction, the methanolic solution was subjected to partition with hexane, yielding the concentrated methanolic phase (CMP) and concentrated hexane phase (CHP) after solvent evaporation in rotatory evaporator. CMP (20.0 g) was fractionated by chromatography on silica gel column using mixtures of hexane, EtOAc and MeOH as eluents in increasing order of polarity. Compound **1** (4.5 mg) was obtained from fraction eluted with hexane/EtOAc (9:1). The fraction

hexane/EtOAc (7:3) was submitted to silica gel column eluted with mixtures of hexane and EtOAc in increasing order of polarity, and the resulting fraction eluted with hexane/EtOAc (8:2) furnished a mixture (30 mg) of compounds **2** and **3**. The CMP fraction eluted with EtOAc (100%) was also fractionated in silica gel column, eluted with mixtures of hexane, EtOAc and MeOH in increasing order of polarity and a mixture (55.7 mg) of compounds **4** and **5** was obtained from the subfraction eluted with hexane/EtOAc (4:6) after washing it with MeOH.

A second extraction from the stem of Salacia impressifolia (2.18 kg) was performed at room temperature successively with hexane and ethyl acetate, and concentrad under vacuum. The ethyl acetate extract (28.0 g) showed particles of a yellow solid. Part of this extract (25.0 g) was washed successively with a solution of hexane/CH₂Cl₂ (1:1) and ethyl acetatel. The fraction washed with hexane/CH₂Cl₂ fractionated 1:1 (3.1)g) was by column chromatography (CC) with silica gel as stationary phase and mixtures of hexane/EtOAc in increasing order of polarity as mobile phase. Subfractions eluted with hexane/EtOAc (9:1) and EtOAc (100%) provided the compounds 6 (40 mg) and 7 (8.5 mg), respectively. The ethyl acetate fraction was also fractionated by column chromatography and from the fractions eluted with hexane/EtOAc (75:25) and hexane/EtOAc (15:85) the compounds 2 and 3 in mixture (1.2 g) and 4 and 5 also in mixture (2.3 g) were obtained, respectively.

RESULTS AND DISCUSSION

Methanolic extract of stem bark from S. impresifolia, after phytochemical procedures, gave one coumarin (1), four triterpenoids (2-5), and two steroids (6 and 7). The structural elucidations of all isolated compounds were performed by means of the comparison of their spectral data (1H, 13C and DEPT NMR) with those ones of the literature. The isolated compounds (1-7) were identified as: isoscopoletin (1) (Waight et al., 1987), quinovic acid (2) (Miana and Alhazimi, 1986), 3-oxo-quinovic acid (3) (Adeoye and Waigh, 1983), $3-O-\beta$ -D-fucopyranosyl quinovic acid (4) (Ferrari *et al.*, 1981), 3-*O*-β-D-quinovopyranosyl quinovic acid (5) (Frou *et al.*, 2003), β -sitosterol (6) and $3-O-\beta$ -D-glucopyranosyl sitosterol (7) (Kojima, 1990). The structures of the compounds isolated are shown in figure 2.





Figure 2: Structures of the compounds 1-7.

The compounds 3-oxoquinovic acid (2), quinovic acid (3), and two glycosylated derivatives of quinovic acid (4) and (5) are not natural products obtained very often. This is the first report on the isolation and identification of the compounds quinovic acid (2), 3oxo-quinovic acid (3), $3-O-\beta$ -D-fucopyranosyl quinovic acid (4) and $3-O-\beta-D$ -quinovopyranosyl quinovic acid (5) from species of Salacia genus. Quinovic acid and its glycosides derivatives quinovic 3β-0-β-Dacid glycopiranoside and quinovic acid-3β-0-β-Dglucopyranosyl-(28 \rightarrow 1)- β -D-glucopyranosyl ester, isolated from *Fagonia cretica*, were reported as having anti-diabetic or DPP-4 inhibiting activity (Saleem et al., 2014). Based on this study, the quinovic acid derivatives 2, 4 and 5 are compounds with potential anti-diabetic activity.

Spectroscopic data of Isoscopoletin (1), Quinovic acid (2) and derivatives (3-5)

Isoscopoletin (1)

Wellow crystals (hexane/EtOAc 9:1) 4.5 mg. ¹H NMR (300 MHz, CDCl₃) δ: 3.95 (3H, *s*, OCH₃-7), 6.27 (1H, *d*, *J*=9.5 Hz, H-3), 6.84 (1H, *s*, H-8), 6.91 (1H, *s*, H-5), 7.60 (1H, *d*, *J*=9,5).

Quinovic acid (2)

White solid (hexane/EtOAc, 8:2) 30.0 mg. ¹³C NMR (75 MHz, Pyridine) δ: 180.0 (C-28), 178.0 (C-27), 134.1 (C-13), 129.0 (C-12), 77.9 (C-3), 56.8 (C-14), 55.7 (C-5), 54.9 (C-18), 48.7 (C-9), 47.2 (C-17), 40.0 (C-8), 39.3 (C-19), 39.2 (C-1), 39.2 (C-4), 37.7 (C-7), 37.5 (C-20), 37.3 (C-10), 37.0 (C-22), 30.5 (C-21), 28.5 (C-15), 28.1 (C-23), 26.3 (C-2), 25.5 (C-16), 23.4 (C-11), 21.3 (C-30), 18.9 (C-6), 18.8 (C-29), 18.2 (C-26), 16.6 (C-25), 16.5 (C-24).

3-oxo-quinovic acid (3)

White solid (hexane/EtOAc, 8:2) 30.0 mg. ¹H NMR (300 MHz, CD₃OD) δ : 0.77-1.23 twelve signals of twelve methyl groups (3H each, *s*, H-23, H-24, H-25, H-26, H-29 and H-30), 6.01 (1H, *s*, H-12). ¹³C NMR (75 MHz,

Pyridine) δ; 216.3 (C-3), 180.0 (C-28), 177.9 (C-27), 134.2 (C-13), 128.6 (C-12), 56.6 (C-14), 54.9 (C-18), 54.7 (C-5), 48.7 (C-17), 47.0 (C-4), 46.2 (C-9), 40.0 (C-7), 39.7 (C-1), 39.3 (C-20), 39.2 (C-8), 37.7 (C-19), 37.0 (C-10), 36.7 (C-22), 34.2 (C-2), 30.5 (C-21), 26.8 (C-23), 25.4 (C-15), 23.4 (C-16), 23.3 (C-11), 21.4 (C-25), 21.3 (C-30), 19.9 (C-6), 18.7 (C-26), 18.2 (C-29), 16.1 (C-24).

3-0- β -D-fucopyranosyl quinovic acid (4)

White solid (hexane/EtOAc, 4:6) 55.7 mg. ¹H NMR (300 MHz, CD₃OD) δ : 0.82-1.00 six signals of six methyl groups (3H each, *s*, H-23, H-24, H-25, H-26, H-29 and H-30), 3.09 (1H, *dd*, *J*=11.5 and 4.3 Hz, H-3), 4.22 (1H, *d*, *J*=7.5 Hz, H-1'), 5.60 (1H, *m*, H-12). ¹³C NMR (75 MHz, CD₃OD) δ : 181.6 (C-28), 179.1 (C-27), 133.8 (C-13), 130.4 (C-12), 107.1 (C-1'), 90.6 (C-3), 75.2 (C-3'), 73.1 (C-2'), 72.9 (C-4'), 71.6 (C-5'), 57.2 (C-14), 56.9 (C-5), 55.5 (C-18), 49.5 (C-17), 48.0 (C-9), 40.6 (C-8), 40.3 (C-4), 40.1 (C-19), 39.9 (C-1), 38.3 (C-20), 38.0 (C-10), 37.8 (C-7), 37.6 (C-22), 31.2 (C-21), 28.5 (C-24), 27.1 (C-2), 26.4 (C-15), 25.7 (C-16), 23.8 (C-11), 21.5 (C-30), 19.2 (C-6), 19.1 (C-23), 18.2 (C-26), 17.1 (C-6'), 16.9 (C-25), 16.9 (C-29).

3-0- β -D-quinovopyranosyl quinovic acid (5)

White solid (hexane/EtOAc, 4:6) 55.7 mg. ¹H NMR (300 MHz, CD₃OD) δ : 0.82-1.00 six signals of six methyl groups (3H each, *s*, H-23, H-24, H-25, H-26, H-29 and H-30), 3.09 (1H, *dd*, *J*=11.5 and 4.3 Hz, H-3), 4.27 (1H, *d*, *J*=7.8 Hz, H-1'), 5.60 (1H, *m*, H-12). ¹³C NMR (75 MHz, CD₃OD) δ : 181.6 (C-28), 179.1 (C-27), 133.8 (C-13), 130.4 (C-12), 106.5 (C-1'), 90.7 (C-3), 77.9 (C-3'), 77.0 (C-5'), 75.8 (C-2'), 72.8 (C-4'), 57.2 (C-14), 56.9 (C-5), 55.5 (C-18), 49.5 (C-17), 48.0 (C-9), 40.6 (C-8), 40.3 (C-4), 40.1 (C-19), 39.9 (C-1), 38.3 (C-20), 38.0 (C-7), 37.8 (C-10), 37.6 (C-22), 31.2 (C-21), 28.5 (C-23), 27.1 (C-2), 26.4 (C-15), 25.7 (C-16), 23.8 (C-11), 21.5 (C-30), 19.2 (C-29), 19.1 (C-6), 18.2 (C-26), 18.2 (C-6'), 17.1 (C-25), 16.9 (C-24).

CONCLUSION

The identification of compounds Quinovic acid (2), 3oxo-quinovic acid (3), 3-O- β -D-fucopyranosyl quinovic acid (4) and 3-O- β -D-quinovopyranosyl quinovic acid (5) belonging to the class of triterpenoids is consistent with the chemical composition of the family Hippocrateaceae and the genus *Salacia*. This is the first report on the isolation and identification of quinovic acid and derivatives from species of the genus *Salacia*. The paper contributes to knowledge of the chemical composition of species, genus and family in study.

ACKNOWLEDGMENT

The authors are grateful to the Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq) and the Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES) for financial support.

REFERENCES

- 1. Adeoye AO and Waigh RD (1983). Secoiridoid and triterpenic acids from the stems of *Nauclea diderrichii*. *Phytochem* 22: 975-978.
- Carvalho PRF, Silva DHS, Bolzani VS and Furlan M (2005). Antioxidant quinonemethide from *Salacia Campestris. Chem Biodivers* 2: 395-398.
- 3. Ferrari F, Cornelio IK, Monache FD and Bettolo GBM (1981). Quinovic acid glycosides from roots of *Macfadyena ungis-cati*. *Planta Med* 43: 24-27.
- 4. Frou A, Tanahashi T, Nagakura N and Nishi T (2003). Two triterpenoid saponins from *Neonauclea sessilifolia*. *Chem Pharm Bul* 51: 1335-1337.
- 5. Kojima H (1990). Sterol glucosides from *Prunella vulgaris*. *Phytochem* 29: 2351-2355.
- 6. Li Y, Huang TH and Yamahara J (2008). *Salacia* root, a unique Ayurvedic medicine, meets multiple targets in diabetes and obesity. *Life Sci* 82: 1045-1049.
- 7. Miana GA and Al-hazimi HMG (1986). Assignment of the 13C NMR spectrum of quinovic acid. *Phytochem* 26: 225-227.
- 8. Morikawa T, Kishi A, Pongpiriyadacha Y, Matsuda H and Yoshikawa M (2003). Structures of new friedelane-type triterpenes and eudesmane-type sesquiterpene and aldose reductase inhibitors from *Salacia chinensis*. *J Nat Prod* 66: 1191–1196.
- 9. Muraoka O, Xie W, Tanabe G, Amer MFA, Minematsu T and Yoshikawa M (2008). On the structure of the bioactive constituents from ayurvedic medicine *Salacia reticulata*: revision of the literature. *Tetrahedron Lett* 49: 7315-7317.
- Nakamura S, Zhang Y, Matsuda H, Ninomiya K, Muraoka O and Yoshikawa M (2011). Chemical structures and hepatoprotective effects of constituents from the leaves of *Salacia chinensis. Chem Pharm Bull.* 59(8): 1020-1028.
- 11. Oe H and Ozaki S (2008). Hypoglycemic effect of 13membered ring thiocyclitol, a novel α -glucosidase inhibitor

CONFLICT OF INTEREST

None declared.

from Kothala-himbutu (*Salacia reticulata*). *Biosci Biotechnol Biochem* 72: 1962-1964.

- 12. Saleem S, Jafri L, ul Haq I, Chang LC, Calderwood D, Green BD, Mirza B (2014). Plants Fagonia cretica L. and Hedera nepalensis K. Koch contain natural compounds with potent dipeptidyl peptidase-4 (DPP-4) inhibitory activity. *J Ethnopharmacol* 156: 26-32.
- 13. Somwong P, Suttisri R and Buakeaw A (2011). A new 1,3diketofriedelane triterpene from *Salacia verrucosa*. *Fitoterapia* 82: 1047–1051.
- 14. Tanabe G, Ogawa W, Xie A, Cao C, Minematsu T, Yoshikawa M and Muraoka O (2009). Facile synthesis of de-O-sulfated salacinols: Revision of the structure of neosalacinol, a potent α -glucosidase inhibitor. *Bioorg Med Chem Let* 19: 2195-2198.
- 15. Thiem DA, Sneden AT, Khan SI and Tekwani BL (2005). Bisnortriterpenes from *Salacia madagascariensis*. *J Nat Prod* 68: 251-254.
- 16. Tinto F, Blair LC, Alli A, Reynolds WF and McLean S (1992). Lupane triterpenoids of *Salacia cordata*. *J Nat Prod* 55: 395-398.
- 17. Waight ES, Razdan TK, Qadri B and Harkar S (1987). Chromones and coumarins from *Skimmia laureola*. *Phytochem* 26: 2063-2069.
- 18. Wang Y, Chen W-S, Wu Z-J, Xi Z-X, Chen W, Zhao G-J, Li X and Sun L-N (2011). Chemical constituents from *Salacia amplifolia*. *Biochem Syst Ecol* 39: 205-208.

Article Citation:

Ripardo Filho HS, Costa NLS, Pacheco PC, Andrade ES, Araújo RNM, Mourão RHV, Guilhon GMSP, Santos LS (2015). Chemical constituents of the stem of *Salacia impressifolia* (Miers) A. C. Smith. Ew J Nat Prod Res 1(1): 1 - 4.